



## Characteristics and Gel-forming Ability of Surimi from Oxeye Scad (*Selar boops*) and Shrimp Scad (*Alepes djedaba*)

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### Abstract

This study aimed to investigate the characteristics and gel properties of surimi prepared from oxeye scad and shrimp scad muscles. The pH and protein solubility of shrimp scad surimi were higher than those of oxeye scad surimi ( $p < 0.05$ ). The  $\text{Ca}^{2+}$ -ATPase activity, sulfhydryl (SH) group and disulfide bond contents were greater in natural actomyosin (NAM) extracted from shrimp scad surimi than oxeye scad surimi ( $p < 0.05$ ). However, kamaboko gel (40°C, 30 min/90°C, 20 min) of oxeye scad surimi had a higher breaking force and deformation ( $p < 0.05$ ) than shrimp scad surimi. Oxeye scad surimi contained larger contents of myoglobin and metmyoglobin resulting in a lowered whiteness of gel compared to shrimp scad surimi ( $p < 0.05$ ). Surimi from oxeye scad showed a superior gel forming ability with an inferior whiteness to shrimp scad. Thus, strategies to improve color of oxeye scad surimi gel should be further studied.

*Keywords: shrimp scad, oxeye scad, gel-forming ability, surimi, physicochemical properties*

### Introduction

Due to the lack of lean fish, which are commonly used for surimi production, dark-fleshed fish have recently received more attention as the raw material for surimi production. In 2009, the catch of pelagic fish in the Gulf of Thailand was approximately 384.9 metric tons (Department of Fisheries 2009). Oxeye scad and shrimp scad are abundant dark-fleshed fish species commonly caught in Southern Thailand, especially in the Thasala coast of Nakhon Si Thammarat. Therefore, the use of these pelagic fish for surimi production is one of a major challenge ways to transform the underutilized fish protein resources into human foods, particularly protein gel-based products. The characteristics of surimi gel depended on the properties of myofibrillar proteins, which were affected by the species and freshness of the fish, as well as on the processing parameters (Niwa 1992). However, no information regarding the characteristics of surimi gel produced from oxeye scad and shrimp scad has been reported. Therefore, the objective of this study was to investigate the characteristics and gel-forming ability of surimi prepared from both species.



## 37 **Materials and Methods**

### 38 **Preparation and chemical analysis of surimi and surimi gel**

39 Oxeye scad with an average weight of 110-120 g and shrimp scad with an average  
40 weight of 125-140 g obtained from the fishing port in Thasala along the coast of the Gulf of  
41 Thailand were used for this study. The fish were washed, filleted manually and minced to  
42 uniformity. Surimi was prepared by conventional washing process using a cold water (4 °C)  
43 /mince ratio of 3:1 (v/w) (Chaijan and others 2004). To prepared the gel, surimi (80%  
44 moisture) was ground with 2.5% (w/w) NaCl to obtain the homogenous sol. The sol was  
45 incubated at 40 °C, 30 min/90 °C, 20 min for kamaboko gel (Chaijan and others 2004).  
46 Modori gel was prepared by incubating the sol at 60 °C, 30 min/90 °C for 20 min (Jiang  
47 2000). The physicochemical properties of surimi and surimi gels were determined. The pH,  
48 myoglobin content, metmyoglobin content and protein solubility were determined by the  
49 method of Benjakul and others (1997), Benjakul and Bauer (2001), Hansen and Sereika  
50 (1969) and Benjakul and Bauer (2000), respectively. Ca<sup>2+</sup>-ATPase activity, reactive SH  
51 group and disulfide bond content of NAM were determined according to the method of  
52 Benjakul and others (1997), Ellman (1959), Thannhauser and others (1987), respectively.  
53 Breaking force and deformation of surimi gel were measured using the texture analyzer  
54 equipped with a spherical plunger (diameter 5 mm, depression speed of 60 mm/min). Surimi  
55 gel color was determined using a colorimeter.  $L^*$ ,  $a^*$  and  $b^*$  were measured and whiteness  
56 was calculated as described by Park (1994).

### 57 **Results and discussion**

58 Oxeye scad muscle contained a larger amount of myoglobin and metmyoglobin when  
59 compared to shrimp scad muscle ( $p<0.05$ ). This indicated that the contents of myoglobin and  
60 metmyoglobin of fish muscle were depended on fish species. The metmyoglobin formation  
61 suggested that myoglobin underwent more oxidation (Benjakul and Bauer 2001). The pH of  
62 shrimp scad surimi was higher than that of oxeye scad surimi (Table 1,  $p<0.05$ ). The  
63 differences in pH between the two species might be due to the differences in chemical  
64 compositions as well as the buffering capacity of muscle. From the results, the lower pH  
65 exhibited the higher relative metmyoglobin content, which was presumably greater in oxeye  
66 scad surimi. Shrimp scad surimi showed a greater protein solubility in 0.6 M KCl than oxeye



67 scad surimi (Table 1). The solubility of a protein is primarily determined by the  
 68 preponderance of charged groups and of hydrophobic groups that are exposed to the solvent  
 69 (Walstra 2003). Oxeye scad surimi might contain the greater proportion of hydrophobic  
 70 groups on the surface of a protein leading to its lowered solubility. A larger proportion of  
 71 charged groups might be found in shrimp scad protein and the solubility was enhanced. The  
 72 Ca<sup>2+</sup>-ATPase activity was greater in NAM extracted from shrimp scad surimi than oxeye  
 73 scad surimi (Table 1,  $p < 0.05$ ). Ca<sup>2+</sup>-ATPase has been used as an indicator of myosin integrity  
 74 (Benjakul and others 1997). Differences protein integrity among fish species were possibly  
 75 caused by different susceptibility to denaturation of muscle protein (Benjakul and others  
 76 2005). Shrimp scad surimi showed greater contents of SH group and disulfide bond compared  
 77 with oxeye scad surimi (Table 1,  $p < 0.05$ ). Benjakul and others (2003) reported that the  
 78 difference in SH content among species was caused by the differences in susceptibility to SH  
 79 oxidation of myofibrillar proteins.

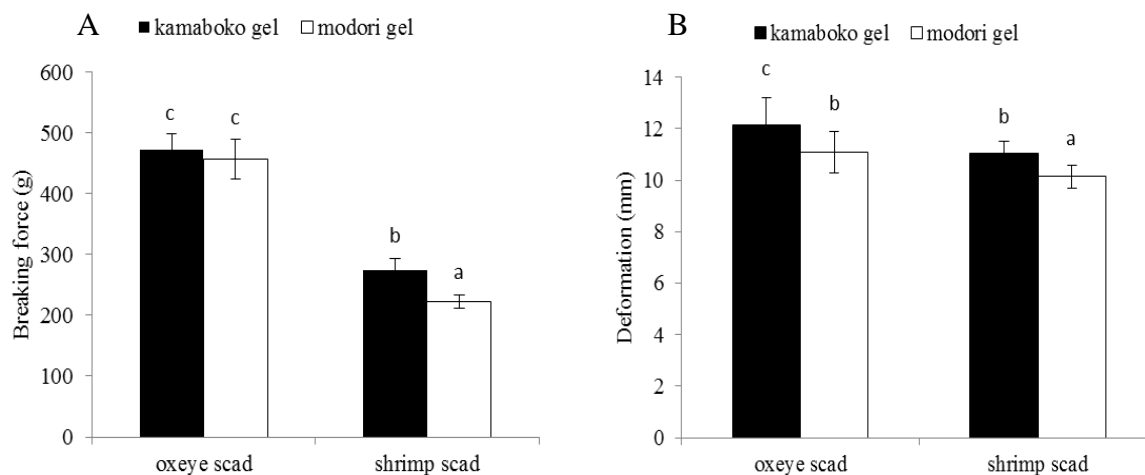
80 **Table 1 Physicochemical properties of surimi from shrimp scad and oxeye scad**

Properties	Shrimp scad	Oxeye scad
Myoglobin (mg/g)	1.28 ± 0.15	2.48 ± 0.11
Relative metmyoglobin (A630/A525)	0.62 ± 0.01	0.71 ± 0.03
pH	6.83 ± 0.04	6.33 ± 0.02
Protein solubility (%)	89.39 ± 1.58	30.55 ± 1.52
Ca <sup>2+</sup> -ATPase (μmol Pi/mg protein/min)	0.19 ± 0.01	0.15 ± 0.01
Sulfhydryl contents (mol/10 <sup>5</sup> g protein)	5.03 ± 0.19	4.31 ± 0.18
Disulfide bond contents (mol/10 <sup>5</sup> g protein)	4.79 ± 0.07	4.16 ± 0.19

81 Values are given as mean ± SD from triplicate determinations.

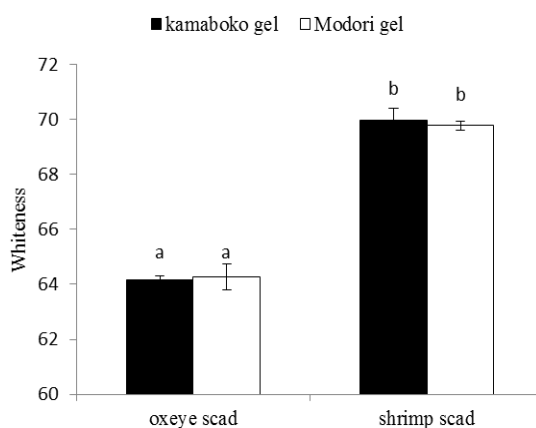
82  
 83 The breaking force and deformation of kamaboko and modori gels from oxeye scad  
 84 surimi were higher than shrimp scad surimi (Figure 1,  $p < 0.05$ ). Protein denaturation was an  
 85 important step for thermal gelation of surimi and it can influence the property of resulting gel.  
 86 Since protein of shrimp scad surimi contains a greater disulfide bond content leading to a  
 87 compact structure of its native conformation. Presumably, the protein of shrimp scad surimi  
 88 underwent partial unfolding upon heating. Protein from oxeye scad surimi contained a lower  
 89 amount of disulfide bond. Thus, it might be susceptible to heat denaturation. Heating of  
 90 protein releases hydrophobic groups and exposures of some buried SH groups (Belitz and  
 91 Grosch 1999). From the result, the more denaturation, the more exposed hydrophobic and SH  
 92 groups can be obtained. As a consequence, hydrophobic interaction and intermolecular  
 93 disulfide bond can be formed in oxeye scad surimi gel to a greater extent resulting in higher  
 94 breaking force and deformation. In addition, the gel strength and elasticity can be influenced

95 by the pH of the surimi. The pH of oxeye scad surimi was closer to the pI of fish myofibrillar  
 96 protein (~5.5) than that of shrimp scad surimi. Thus, a more protein-protein interaction can be  
 97 taken place in oxeye scad surimi gel.



98  
 99 **Figure 1** Breaking force (A) and deformation (B) of surimi gels prepared from oxeye scad and  
 100 shrimp scad. Bars represent the standard deviation from seven determinations for  
 101 breaking force and deformation. Different letters indicate significant differences ( $p < 0.05$ ).

102 Gels from shrimp scad generally showed a higher whiteness than those from oxeye  
 103 scad (Figure 2,  $p < 0.05$ ). Oxeye scad surimi contained larger contents of myoglobin and  
 104 metmyoglobin resulting in a lowered whiteness of gel compared to shrimp scad surimi. Due  
 105 to the higher content of myoglobin in oxeye scad surimi, the greater oxidation might take  
 106 place in this species, compared with shrimp scad. Metmyoglobin formation, caused by  
 107 oxidation process, possibly resulted in the darker color of oxeye scad surimi gel.



108  
 109 **Figure 2** Whiteness of surimi gels prepared from oxeye scad and shrimp scad. Bars represent the  
 110 standard deviation from triplicate determinations for breaking force and deformation.  
 111 Different letters indicate significant differences ( $p < 0.05$ ).

112 **Conclusion**

113           The physicochemical properties of surimi gel were species-dependent. Oxeye scad  
114 surimi gel exhibited the higher breaking force and deformation with lower whiteness than  
115 that of shrimp scad surimi gel.

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